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9.1 Summary

9.1.1 Biological samples are made acidic with monosodium phosphate buffer (pH 5.1) and extracted with a mixture of n-butyl chloride and ethyl ether. The extract is derivatized with trimethylsulfonium hydroxide and an aliquot is injected into a GC equipped with an NPD detector for quantitation. The aliquot is subsequently confirmed using gas chromatography mass spectrometry.

9.2 Specimen Requirements

9.2.1 1 mL of fluid(s) or 1 g of tissue(s) or comparable amounts of fluid or tissue dilutions/homogenates

9.3 Reagents and Standards

- 9.3.1 Butalbital, 1 mg/mL
- 9.3.2 Pentobarbital, 1 mg/mL
- 9.3.3 Secobarbital, 1 mg/mL
- 9.3.4 Phenobarbital, 1 mg/mL
- 9.3.5 Amobarbital, 1 mg/mL
- 9.3.6 Butabarbital, 1 mg/mL
- 9.3.7 Glutethamide, 1 mg/mL
- 9.3.8 Phenytoin (diphenylhydantoin), 1 mg/mL
- 9.3.9 Carbamazepine, 1 mg/mL
- 9.3.10 Cyclopentalbarbital (cyclopal), internal standard
- 9.3.11 Trimethyl sulfonium iodide
- 9.3.12 Monosodium phosphate
- 9.3.13 Silver oxide
- 9.3.14 Hexane
- 9.3.15 Isoamyl alcohol
- 9.3.16 Methanol
- 9.3.17 Toluene
- 9.3.18 N-butyl chloride
- 9.3.19 Diethyl ether

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- 9.4 Solutions, Internal Standard, Calibrators, Controls
 - 9.4.1 1.5 M monosodium phosphate buffer, pH 5.1 Add 103.4 grams of monosodium phosphate (NaH₂PO₄) to a 500 mL volumetric flask and QS to volume with dH₂O.
 - 9.4.2 n-Butyl chloride:diethyl ether (95:5, v:v) Mix 950 mL n-butyl chloride with 50 mL diethyl ether .
 - 9.4.3 Toluene:Hexane:Isoamyl Alcohol (THIA) (78:20:2, v:v:v) Mix 78 mL toluene, 20 mL hexane and 2 mL isoamyl alcohol.
 - 9.4.4 Reconstituting solvent: Toluene/hexane/isoamyl alcohol/methanol, (59:15:1.5:25, v:v:v:v): Mix 25 mL methanol with 75 mL THIA
 - 9.4.5 Trimethyl sulfonium hydroxide derivatizing reagent Add 6.12 g trimethylsulfonium iodide, 7.39 g silver oxide, and 15 mL methanol to a 25 mL teflon capped test tube covered with aluminum foil (light sensitive reaction). Rotate for four or more hours, centrifuge, and decant the supernatant to an aluminum foil covered test tube. Keep refrigerated.
 - 9.4.6 Extraction Solvent containing Internal Standard: Weigh 20 mg of cyclopal free acid and transfer to a 10 mL volumetric flask. QS to volume with methanol for final concentration of 2 mg/mL). Aliquot 2 mL of 2 mg/mL cyclopentabarbital (cyclopal) stock solution into a 1000 mL volumetric flask and QS to volume with extraction solvent (n-butyl chloride:diethyl ether) to yield 4 mg/L cyclopal in extraction solvent.
 - 9.4.7 Drug stock solutions:
 - 9.4.7.1 If 1 mg/mL commercially prepared stock solutions are not available, prepare 1 mg/mL solutions from powders. Weigh 10 mg of the free acid, transfer to a 10 mL volumetric flask and QS to volume with methanol. Note: If using the salt form, determine the amount of the salt needed to equal 10 mg of the free acid, and weigh this amount.
 - 9.4.8 Working Standard Solution A (0.1 mg/mL): Add 1.0 ml of each of the following 1 mg/mL stock solutions to a 10 mL volumetric flask: butalbital, phenobarbital, carbamazepine and phenytoin. QS to volume with methanol.
 - 9.4.9 Working Standard Solution B (0.1 mg/mL): Add 1.0 ml of each of the following stock solutions to a 10 mL volumetric flask: butabarbital, secobarbital, pentobarbital, and glutethamide. QS to volume with methanol.
 - 9.4.10 Blood calibrators, standards, and controls preparation:
 - 9.4.10.1 To prepare the following calibration curve, pipet the following volumes of working standard solution A into appropriately labeled 16 x 125 mm screw cap test tubes

| • | 30 mg/L Calibrator | 300 µL of working standard solution A |
|---|--------------------|---------------------------------------|
| • | 20 mg/L Calibrator | 200 µL of working standard solution A |
| • | 10 mg/L Calibrator | 100 μL of working standard solution A |
| • | 5 mg/L Calibrator | 50 μL of working standard solution A |
| • | 2 mg/L Calibrator | 20 μL of working standard solution A |
| • | 1 mg/L Calibrator | 10 µL of working standard solution A |

- 9.4.10.1.1 Evaporate standards to dryness under nitrogen. Add 1 mL blank blood to each tube.
- 9.4.10.2 Standard B contains rarely encountered drugs (butabarbital, secobarbital, pentobarbital, and glutethamide).

 During routine barbiturate analyses, run at least 1 standard containing working solution B for retention times.

 If any of the 4 drugs are present, a full calibration curve is required.

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| | | | 9.4.10.2.1 | screw-cap to | | | rd Solution B into a 16 x 125 mm labeled ogen. Add 1 mL blank blood for a final |
| | | | 9.4.10.2.2 | | bration curve is require opriately labeled 16 x 1 | | ing volumes of working standard solution test tubes |
| | | | | 20 mg/I 10 mg/I 5 mg/L 2 mg/L | Calibrator Calibrator Calibrator Calibrator Calibrator | 200 μL of working 100 μL of working 50 μL of working 20 μL of working 10 μL of working | g standard solution B r nitrogen. Add 1 mL blank blood to each |
| | | 9.4.10.3 | Controls | | | | |
| | | | 9.4.10.3.1 | | ontrol. Blood bank blood carbamazepine and glu | | determined not to contain barbiturates, |
| | | | 9.4.10.3.2 | | ntrol. In house control nanufacturer than stand | • | alyte of interest from a different lot |
| 9.5 | Appara | atus | | | | | |
| | 9.5.1 | Agilent GC with Nitrogen-Phosphorous Detector, Chemstation software, compatible computer & printer Test tubes, 16 x 125 mm round bottom, screw cap tubes, borosilicate glass with Teflon caps | | | | | |
| | 9.5.2 | | | | | | |
| | 9.5.3 | | | | | | |
| | 9.5.4 | | | | | | |
| | 9.5.5 | Centrifug | ge capable of | 2,000 – 3,00 | 0 rpm | | |
| | 9.5.6 | Vortex m | nixer | | | | |
| | 9.5.7 | Evaporat | or/concentrat | tor | | | |
| | 9.5.8 | GC autos | sampler vials | and inserts | | | |
| | 9.5.9 | Test tube | rotator | | | | |
| | 9.5.10 | GC/NPD | parameters. | Instrument c | conditions may be cha | nged to permit im | proved performance. |
| | | 9.5.10.1 | Oven progr | ram. | | | |
| | | | EquilibreInitial toInitial tiRamp:Final To | me: | 0.50 minutes 150° C 0.5 minutes 12° C/min 280° C | | |

• Final Temp:

 $280^{\circ}\,\mathrm{C}$

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Final Time: 4 minutesRun Time: 15 minutes

9.5.10.2 Inlet.

Mode: Splitless
Temperature: 250° C
Constant pressure: 16 psi
Purge flow: 49.6 mL/min
Total flow: 52.9 mL/min
Injection volume: 1.0 μL

9.5.10.3 Detector.

Temperature: 290° C
Hydrogen flow: 3.0. mL/min
Air flow: 60 mL/min

• Mode: Constant column + makeup flow

Combined flow: 20.0 mL/min
 Injection volume: 1.0 μL
 Makeup flow: On

9.5.10.4 Column: HP-5 25 m x 0.25 mm x 0.25 μm.

9.5.11 GC/MSD parameters. Instrument conditions may be changed to permit improved performance.

9.5.11.1 Acquisition Mode: Scan (50 – 550 amu)

9.5.11.2 Column: HP 5MS 25 m x 0.25 mm x 0.25 μ m

9.5.11.3 Detector Temperature: 280° C

9.5.11.4 Basic drug screen. Instrument conditions may be changed to permit improved performance.

9.5.11.4.1 Oven Program

Equilibration time: 0.50 minutes
Initial temp: 110° C
Initial time: 1 minutes
Ramp: 10° C/min
Final Temp: 290° C
Final Time: 9 minutes
Run Time: 28 minutes

9.5.11.4.2 Inlet

Mode: Splitless
 Temperature: 270° C
 Injection volume: 1.0 μL

• Purge Time: ON at 1.0 minute

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| 9.6 Procedure | |
| 9.6.1 Label clean 16 x 125 mm screw cap tubes accordingly, negative, calib | rators, control(s) and case sample IDs. |

- 9.6.2 Prepare calibrators and controls.
- 9.6.3 Pipet 1 mL of each case sample into appropriately labeled tubes.
- 9.6.4 Add 1 mL 1.5 M sodium phosphate buffer (pH 5.1) to each tube.
- 9.6.5 Add 3 mL extraction solvent (n-butyl chloride/diethyl ether) containing cyclopal internal standard to each tube.
- 9.6.6 Cap and rotate tubes for 30 minutes.
- 9.6.7 Centrifuge at approx 2500 rpm for 10 minutes. Transfer organic (upper) layer to clean 5 mL conical bottom tubes. Discard lower layers.
- 9.6.8 Add 50 μ L methanol and 50 μ L TMSH to each tube.
- 9.6.9 Evaporate samples to dryness under nitrogen at 50-55° C. Note: Do not evaporate above 60° C.
- 9.6.10 Reconstitute samples with 1.0 mL toluene/hexane/isoamyl alcohol/methanol reconstituting solvent. Vortex briefly.
- 9.6.11 Transfer small aliquot to appropriately labeled GC vials and inject 1-2 µl on GC-NPD.
- 9.6.12 Save remainder of reconstituted samples for confirmation by GC-MSD (if not already confirmed).

9.7 Calculation

9.7.1 Calculate the concentrations by interpolation of a linear plot of the response curve based on peak height (or area) ratios versus calibrator concentration.

9.8 Quality Control And Reporting

9.8.1 See Toxicology Quality Guidelines

9.9 References